

Development and validation of RP-HPLC methods for estimation of Emtricitabine

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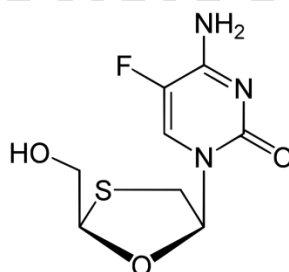
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Abstract : High-performance liquid chromatographic (HPLC) was developed and validated for the quantitative determination of Emtricitabine. The RP-HPLC method was developed by the isocratic technique on a reversed-phase, Phenomenex column. The retention time for EC was 5.9 min, this method can be a cheap, reliable and less time consuming chromatographic analysis. The proposed method is highly sensitive, precise and accurate and hence successfully applied for determining the assay of a marketed formulation.

Key Words: Emtricitabin, Correlation equation, retention time, RP-HPLC.

INTRODUCTION

Emtricitabine is a nucleoside analogue and reverse transcriptase inhibitor used for the treatment and prevention of human immunodeficiency virus (HIV) infection and the acquired immunodeficiency syndrome (AIDS). Its chemical name is 4-amino-5-fluoro-1-[(2R,5S)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl]-1,2-dihydropyrimidine-2-one. A fixed-dose triple combination of Emtricitabine, Tenofovir and efavirenz (Sustiva, marketed by Bristol-Myers Squibb) was approved by the U.S. Food and Drug Administration (FDA) on July 12, 2006 under the brand name Atripla. Emtricitabine/Tenofovir was approved for medical use in the United States in 2004 (WHO, 2019).



Literature survey reveals stability indicating UV spectrophotometric, RP-HPLC (Gerber, et.al., 2014, Morgan, et.al., 2015, Karger, et.al., 1997, Henry, et.al., 2009). HPLC in human serum, HPTLC (Geiss, et.al., 1987, Jork, et.al., 1990, Hahn, et.al., 2000) and LC-MS (Chaimbault, et.al., 2014, Dass, et.al., 2007, Pitt, et.al., 2017 Niessen, et.al., 2006) method for the estimation of EC. Several analytical methods have been reported for the quantitative determination of EC such as high performance liquid chromatography, HPTLC- densitometry and LC-MS. A new analytical method needs to be established. In the present work we have developed a simple UV method for estimation of EC correlation equation; obtained from absorbance maxima of EC. The values obtained from the developed equation were also verified using HPLC. The existing HPLC method was further modified to reduce the retention time. Both these methods are used to calculate drug content in marketed formulation and in dissolution media.

RESULTS AND DISCUSSION

MATERIALS AND METHODS :

EMTRICITABINE :

The Reverse Phase-High Performance Liquid Chromatography (RP-HPLC) was carried out on Waters Alliance Series with 2695 Separations Module and 2998 Photodiode Array Detector using Empower 2 software.

ANALYTICAL CONDITIONS

The active ingredient content of Emtricitabine was determined by HPLC analysis. The following HPLC conditions were used in the analysis.

Mobile phase	Column	Column Specifications	Column Temperature	Flow Rate	Injection Volume	Detector	Detector Wave length	Retention Time
Acetonitrile : HPLC Water adjusted pH 4.0 with Orthophosphoric acid (75 : 25% v/v)	Phenomenex	250 mm x 4.6 mm, 5µm	30 °C	1.0 mL/min	10µL	PDA	230nm	5.9min (approximately)

Table 1

RECOMMENDED PROCEDURES:

After systematic and detailed study of the various parameters involved, as described under results and discussion in this paper, the following procedure was recommended for the determination of ECB in bulk samples and pharmaceutical formulations.

PREPARATION OF SOLUTIONS:**PREPARATION OF STANDARD SOLUTION:**

0.01840 g [18.40 mg] of Emtricitabin standard was weighed accurately into a clean and dry 25 ml volumetric flasks and dissolved with 5 ml of Acetonitrile (HPLC grade) and made upto the mark with the same solvent. These solutions were marked as R1A respectively.

PREPARATION OF SAMPLE SOLUTION:

0.01820 g [18.20mg], 0.02030 g [20.30mg], 0.01860 g [18.60mg], 0.01850 g [18.50 mg] and 0.01880 g [18.80mg] of Emtricitabine was weighed accurately into a clean and dry five separate 25 ml volumetric flasks and dissolved with 10 ml of Acetonitrile (HPLC grade) respectively and made upto the mark with same solvent.

PREPARATION OF LINEARITY SOLUTIONS:

0.03320 g [33.20 mg] of Emtricitabine standard (99.5%) was weighed accurately into a clean and dry 10 ml volumetric flask and dissolved with 5 ml acetonitrile (HPLC grade) and made up to the mark with the same solvent. This prepared solution is equivalent to 3303 mg/L. Lower concentrations were made by appropriate dilutions as mentioned in the table 2.22.

Concentration of solution taken for dilution (µg/ml)	Dilution details		Concentration of diluted solution (µg/ml)
	Volume taken (ml)	Volume made up (ml)	
3303	7.569	10	2500
2500	8.000	10	2000
2000	7.500	10	1500
1500	6.667	10	1000
1000	5.000	10	500

Table 2.22.: Dilution details linearity solutions

PREPARATION OF FORTIFICATION STANDARD SOLUTION 0.01850 g [18.50 mg] and 0.01840 g [18.40 mg] of Emtricitabine standard was weighed accurately into two separate clean and dry 25 ml volumetric flasks and dissolved with 10 mL of acetonitrile (HPLC grade) and made upto the mark with the same solvent. These solutions were marked as R1A and R1B respectively

PREPARATION OF 5 G/KG FORTIFICATION LEVEL SOLUTION

0.0186 g [18.6 mg] of Emtricitabine technical (test item) was weighed accurately into a clean and dry 25 ml volumetric flask, 0.093 ml (93 µL) of linearity prepared solution of (1000 µg/ml) was added to it and dissolved with acetonitrile (HPLC grade) and made upto the mark with the same solvent. This solution marked as T1 was equivalent to dose concentration (5 g/kg).

PREPARATION OF 10 G/KG FORTIFICATION LEVEL SOLUTION

0.0186 g [18.6 mg] of Emtricitabine technical (test item) was weighed accurately into a clean and dry 25 ml volumetric flask, 0.186 ml (186 µL) of linearity prepared solution of (1000 µg/ml) was added to it and dissolved with acetonitrile (HPLC grade) and made upto the mark with the same solvent. This solution marked as T2 was equivalent to dose concentration (10 g/kg).

DETERMINATION OF ACTIVE INGREDIENT CONTENT**PREPARATION OF STANDARD SOLUTION OF EMTRICITABIN**

0.01850 g [18.50 mg] and 0.01840 g [18.40 mg] of Emtricitabine standard was weighed accurately into two separate clean and dry 25 ml volumetric flasks and dissolved with 10 ml of acetonitrile (HPLC grade) and made upto the mark with the same solvent. These solutions were marked as R1A and R1B respectively.

PREPARATION OF SAMPLE SOLUTION FOR ACTIVE INGREDIENT

0.01820 g [18.20 mg] and 0.01850 g [18.50 mg] of Emtricitabine TC was weighed accurately into two different clean and dry 25 ml volumetric flasks and dissolved with 10 ml of acetonitrile (HPLC grade) and made up to the mark with same solvent. These solutions were marked as R2A and R2B respectively.

LINEARITY

Sample code	Standard CS1	Standard CS2	Standard CS3	Standard CS4	Standard CS5	Slope	Correlation Co-efficient	Intercept
Conc. (µg/ml)	500	1000	1500	2000	2500			
Observed Area	6971035	12174534	17394224	22615938	27636329	10354	0.999	1826814

Table No.2.23: Results of linearity.

Each of these solutions (2500, 2000, 1500 ,1000 and 500 mg/L) were injected into HPLC under the given conditions and the peak area was recorded and a graph of detector response (peak area) versus concentration in mg/L was plotted. The values for slope (m), intercept (b) and the linear regression coefficient (R²) were calculated.

The detector response to varying concentrations of Emtricitabin was found to be linear (R² = 0.9998) in the range of 500 to 2500 mg/L. The plot of concentrations versus detector response along with the regression parameters was attached and the results are presented in Table. No. 2.23

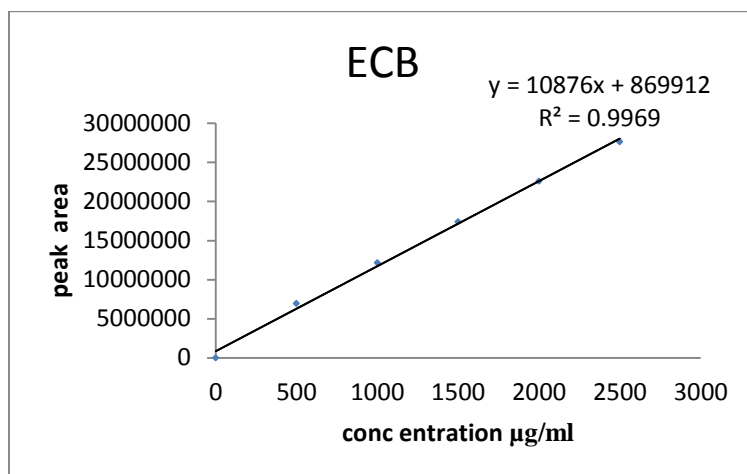


Fig. Linearity curve of Emtricitabin reference item

ACTIVE INGREDIENT CONTENT

The active ingredient is the drug substance and excipients which are ingredients of the tablet. It has the property of long term stabilization, and bulking of up solid formulation which contains small amount to a therapeutic enhancement. The above prepared Emtricitabine standard solutions and sample solutions were injected into a High performance liquid chromatography (HPLC) by using the analytical conditions. Emtricitabine active ingredient content, present in the sample was quantified by injecting reference standard and comparing the peak area of standard with the peak area of sample. The results are presented in table no.2.26.

Sample Code	Standard Wt (mg)	Standard Purity (g/kg)	Standard Area	Mean Standard Area	Sample Area	Sample Wt (grams)	A.I. Content (g/kg)	A.I. Content (% w/w)	Mean A.I. Content (% w/w)		
R1A1	18.50	995.00	8686035	8686715	8384660	18.20	976.23	97.62	97.70		
R2A1										8397127	977.68
R2A2											
R1B1	18.40		8678334								
R1A2	18.50		8687395								
R2B1			8677025	8559508	18.50	976.22	97.62				
R2B2						8566189	976.98	97.70			
R1B2	18.40		8675715								
Mean										97.68	
Standard deviation										0.03	
Relative standard deviation (%RSD)									0.03		

Table 2.26 Results of Active Ingredient:

CONCLUSIONS

Emtricitabine was evaluated as per the guidelines of ICH. The method was validated for the determination of Active Ingredient Content in EC for the test item EC meets the acceptance criteria. The active ingredient content of the test item EC was found to be 97.68(% w/w) by using HPLC method.

REFERENCES

- Chaimbault, Patrick. 2014. "The Modern Art of Identification of Natural Substances in Whole Plants". In Jacob, Claus; Kirsch, Gilbert; Slusarenko, Alan; Winyard, Paul G.; Burkholz, Torsten (eds.). Recent Advances in Redox Active Plant and Microbial Products. Springer Netherlands. pp. 31–94. doi:10.1007/978-94-017-8953-0_3. ISBN 9789401789523.
- Dass, Chhabil (2007-01-01). "Hyphenated Separation Techniques". Fundamentals of Contemporary Mass Spectrometry. John Wiley & Sons, Inc. pp. 151–194. doi:10.1002/9780470118498.ch5. ISBN 9780470118498.
- Geiss, F.1987. Fundamentals of thin layer chromatography planar chromatography, Heidelberg, Hüthig, ISBN 3-7785-0854-7

- Gerber, F, Krummen, M, Potgeter, H, Roth, A, Siffrin, C.; Spoendlin, C. 2004. "Practical aspects of fast reversed-phase high-performance liquid chromatography using 3µm particle packed columns and monolithic columns in pharmaceutical development and production working under current good manufacturing practice". *Journal of Chromatography A*. 1036 (2): 127–133. doi:10.1016/j.chroma.2004.02.056. PMID 15146913.
- Hahn-Deinstorp, E. 2000. *Applied Thin-Layer Chromatography. Best Practice and Avoidance of Mistakes*, Wiley-VCH, Weinheim, ISBN 3-527-29839-8
- Henry, Richard A. 2009 "The Early Days of HPLC at Dupont". *Chromatography Online*. Avanstar Communications Inc.
- Jork, H., Funk, W., Fischer, W., Wimmer, H. 1990. *Thin-Layer Chromatography: Reagents and Detection Methods, Volume 1a*, VCH, Weinheim, ISBN 3-527-27834-6
- Jork, H., Funk, W., Fischer, W., Wimmer, H. 1994. *Thin-Layer Chromatography: Reagents and Detection Methods, Volume 1b*, VCH, Weinheim
- Karger, Barry L. 1997. "HPLC: Early and Recent Perspectives". *Journal of Chemical Education*. 74 (1): 45. Bibcode:1997JChEd..74...45K. doi:10.1021/ed074p45.
- Morgan, David J. 2003-11-19. "Fraction collector (post on Flickr)". Flickr. Retrieved 28 October 2015.
- Niessen, Wilfried M. A (2006). *Liquid Chromatography-Mass Spectrometry, Third Edition*. Boca Raton: CRC Taylor & Francis. pp. 50–90. ISBN 9780824740825. OCLC 232370223.
- Pitt, James J (2017-03-12). "Principles and Applications of Liquid Chromatography-Mass Spectrometry in Clinical Biochemistry". *The Clinical Biochemist Reviews*. 30 (1): 19–34. ISSN 0159-8090. PMC 2643089. PMID 19224008.
- World Health Organization. 2019. *World Health Organization model list of essential medicines: 21st list 2019*. Geneva: World Health Organization. hdl:10665/325771. WHO/MVP/EMP/IAU/2019.06. License: CC BY-NC-SA 3.0 IGO.

